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BRIEF REPORT

Presence of antibodies against *Leptospira* serovars in *Chaetophractus villosus* (Mammalia, Dasypodidae), La Pampa province, Argentina

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KEYWORDS

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Abstract Leptospirosis is a zoonosis of worldwide distribution. The aim of this study was to examine the presence of antibodies against 21 *Leptospira* reactive serovars in *Chaetophractus villosus* in La Pampa province, Argentina, using the microscopic agglutination test (MAT). Pathologic changes compatible with leptospirosis and *in situ* detection of the agent by immunohistochemistry were studied in 24 and 3 individuals respectively. Only 35/150 (23.3%) serum samples had antibodies against *Leptospira* sp. Six percent of the samples reacted with serovar Canicola, 4.7% with serovar Castellonis, 1.3% with serovar Icterohemorrhagiae and 0.7% with serovar Hardjo. Sixteen (10.6%) serum samples agglutinated with Castellonis–Icterohemorrhagiae and Canicola–Castellonis serovars, both with 4.7%, and Canicola–Hardjo and Castellonis–Canicola–Icterohemorrhagiae both with 0.6%. Fourteen animals had variable degrees of lesions, which were more severe in animals with higher serological titers (3200), and *Leptospira* sp. was detected in 3 animals by immunohistochemistry. These results represent the first record of the presence of *Leptospira* in *C. villosus* in La Pampa. © 2015 Asociación Argentina de Microbiología. Published by Elsevier España, S.L.U. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

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PALABRAS CLAVE

Leptospira;
Serovares;
Chaetopractus villosus;
Animales silvestres;
Xenarthra

Presencia de anticuerpos contra serovares de *Leptospira* en *Chaetopractus villosus* (Mammalia, Dasypodidae) en la provincia de La Pampa, Argentina

Resumen La leptospirosis es una zoonosis de distribución mundial. Nuestro objetivo fue examinar la presencia de anticuerpos contra 21 serovares reactivos de *Leptospira* en *Chaetopractus villosus* en la provincia de La Pampa, Argentina, mediante la prueba de aglutinación microscópica (MAT). Se realizó el estudio histopatológico y la detección *in situ* del agente por inmunohistoquímica en 24 y 3 individuos, respectivamente. Solo 35/150 (23,3%) muestras de suero presentaron anticuerpos contra *Leptospira* sp. Seis por ciento reaccionaron al serovar Canicola; 4,7% a Castellonis; 1,3% a Icterohemorrhagiae y 0,7% a Hardjo. Dieciséis (10,6%) sueros aglutinaron con Canicola-Castellonis y Castellonis-Icterohemorrhagiae, ambos con 4,7%, y con Canicola-Hardjo y Castellonis-Canicola-Icterohemorrhagiae, ambos con 0,6%. En 14 animales se encontraron lesiones compatibles, las que resultaron más graves en animales con títulos serológicos elevados (3200). En 3 animales estudiados se detectó el agente causal por inmunohistoquímica. Estos resultados constituyen los primeros registros de la presencia de *Leptospira* en *C. villosus* en La Pampa.

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Leptospirosis is an infectious disease of great public health concern. It is considered the zoonosis with the largest global distribution¹. This disease is generally caused by two major pathogenic genomospecies: *Leptospira borgpetersenii* and *Leptospira interrogans sensu stricto*³. Each serovar has one or more preferred host animals; however, each animal species can be a host to one or more serovars, whereas humans can be hosts to many serovars.

Domestic and wild animals are important reservoirs of *Leptospira*, which is always excreted through the urine in a discontinuous way and in varying periods of time. The transmission to humans is the result of exposure to the urine of infected animals by direct contact or through the water. The rural areas are at a greater risk because of the manipulation of domestic and wild animals for meat consumption; armadillos (*Xenarthra*, *Dasypodidae*) are a clear example of this situation. *Chaetopractus villosus* is a member of the superorder *Xenarthra*, and its distribution range extends from the arid Gran Chaco region, which is located between Bolivia, Paraguay and northern Argentina, to as far south as the Argentine Tierra del Fuego and Magallanes in Chile⁵. *C. villosus* is an omnivore that feeds on insects, invertebrates, small vertebrates, seeds and carrion (infected animal tissues such as fetuses and placentas) of infected animals. *Leptospiras* in *C. villosus* have been detected only in Buenos Aires province, Argentina. Different serovars have been isolated, including Paidjan, Argentinensis, Hardjo, Canicola, Bataviae and *Leptospira biflexa*^{2,4,8,13,14}. In addition, the following serovars were identified by serology: Hardjo, Wolffi, Paidjan, Argentinensis, Bataviae, Canicola, Sejroe, Hebdomadis, Pomona, Castellonis, Grippotyphosa and Icterohaemorrhagiae^{4,8}.

No updated information has been published since the end of the 70s, and no publications are available from La Pampa province about *Leptospira* in *C. villosus* populations, except for the province of Buenos Aires^{10,11}.

For this reason, the aim of this study is to know the seroprevalence of antibodies against *Leptospira* serovars in *C. villosus* from La Pampa province, Argentina, through the microscopic agglutination test (MAT) and to describe the presence of histopathological lesions compatible with the detection of *Leptospira* by immunohistochemistry.

The capture site is located in central La Pampa, where the weather is characterized by hot, rainy summers with temperatures over 35 °C and cold winters with average temperatures of 10 °C, including frequent and severe ground frosts. Rains are common in spring and autumn, with winter being the driest season. Annual rainfall varies from 450 mm to 800 mm. Beef is the most important production in the region. Management is extensive and cows are free-range year round, with stocking rates depending on the available pasture, which in turn depends on the season and weather conditions. Cattle stocking rates have an average of 0.75 cows per hectare.

C. villosus were captured with permission of the Ministry of Production, Secretariat of Agricultural and Natural Resources Directorate of La Pampa province, and the agreement of farm owners. The armadillos were trapped and carried to the laboratory under adequate care. In order to determine the age, the specimens were measured (from snout to tail tip), and their general appearance was evaluated. Armadillos with a length equal to or less than 460 mm were considered juveniles, whereas longer specimens were classified as adults.

Blood was extracted from the caudal vein of 150 armadillos captured between 2007 and 2010 in La Pampa province. The blood samples were centrifuged for 15 min at 2500 rpm. Sera were separated and stored at -20 °C until the time of analysis and testing for the presence of *Leptospira* sp. antibodies. The MAT was used in the analysis of antibodies against *Leptospira* sp., using the following serovars as antigens: Castellonis, Canicola, Celledoni, Icterohaemorrhagiae, Hardjo, Pomona, Grippotyphosa, Pyrogenes,

Ranarum, Hebdomadis, Sarmin, Bataviae, Mini, Autumnalis, Cynopteri, Panama, Australis, Javanica, Djasiman, Wolffi and Tarassovi. Initial serum dilution was 1:25, and sera with a titer of 50 or higher were considered positive.

Twenty four *C. villosus*, with previous serological tests for *Leptospira* (14 positive and 10 negative) were euthanized under anesthesia (tiletamine and zolazepam, 5.0 mg/kg/I.M), respecting the guidelines of the Canadian Council of Animal Care referring to working with experimental animals used in scientific research. Kidneys were extracted for renal histology in order to look for lesions compatible with leptospirosis and fixed in 10% formaldehyde solution. Then, kidney samples were embedded in paraffin, sectioned and stained with hematoxylin-eosin. Finally, three samples with lesions compatible with leptospirosis infection were used for *in situ* detection of *Leptospira* by immunohistochemistry following already described procedures⁵. Rabbit polyvalent antibody reactive against serovar Canicola, Pomona and Icterohaemorrhagiae (USDA) was used as conjugate and positive results were revealed with AEC (aminoethylcarbazole solution) as chromogenic substrate. Finally samples were stained with hematoxylin.

The sacrificed animals were deposited in the mammalian collection of the Universidad Nacional de La Pampa (UNLPam MA under the following numbers: UNLPam MA609; UNLPam MA639; UNLPam MA651; UNLPam MA652; UNLPam MA653; UNLPam MA660; UNLPam MA661; UNLPam MA664; UNLPam MA673; UNLPam MA676; UNLPam MA697; UNLPam MA698; UNLPam MA703; UNLPam MA704; UNLPam MA706; UNLPam MA709; UNLPam MA727; UNLPam MA739; UNLPam MA742; UNLPam MA744; UNLPam MA773; UNLPam MA776; UNLPam MA786; UNLPam MA788; UNLPam MA789; UNLPam MA793; UNLPam MA795; UNLPam MA799; UNLPam MA800; UNLPam MA803; UNLPam MA810; UNLPam MA815; UNLPam MA825; UNLPam MA836; UNLPam MA843). It is worth noting that *C. villosus* is not an endangered species⁹.

Statistical analyses were performed using the Chi-square test for two variables: age and sex. Statistical significance in this study was defined at the $p \leq 0.05$ levels (Epi Info 6.0.4 software).

Out of the 150 *C. villosus* analyzed, 35 (23.3%) had antibodies against *Leptospira* (95% confidence intervals 16.5–30.2, Table 1).

Seventy of the 150 samples corresponded to male specimens, 18 (25.7%) of which were positive for *Leptospira*, whereas the remaining 80 samples were obtained from females, 17 (21.3%) of which showed positive results. Based on their length, 127 of the *C. villosus* examined were adults, 34 (26.7%) of which had positive results for *Leptospira*, whereas only 1/23 (4.3%) juveniles showed positive results.

Of the 150 serum samples tested, 6% resulted positive to serovar Canicola, 4.7% to Castellonis, 1.3% to Icterohaemorrhagiae and 0.7% to Hardjo. Finally, 10.6% of the serum samples agglutinated with two or more serovars, being the Castellonis–Icterohaemorrhagiae and Canicola–Castellonis serovars the most frequently observed patterns, both with 4.7%, and Canicola–Hardjo and Castellonis–Canicola–Icterohaemorrhagiae with 0.6%. The highest titers were observed with serovars Canicola, Castellonis (3200) and Icterohaemorrhagiae (800). No antibodies were detected against serovars Grippotyphosa,

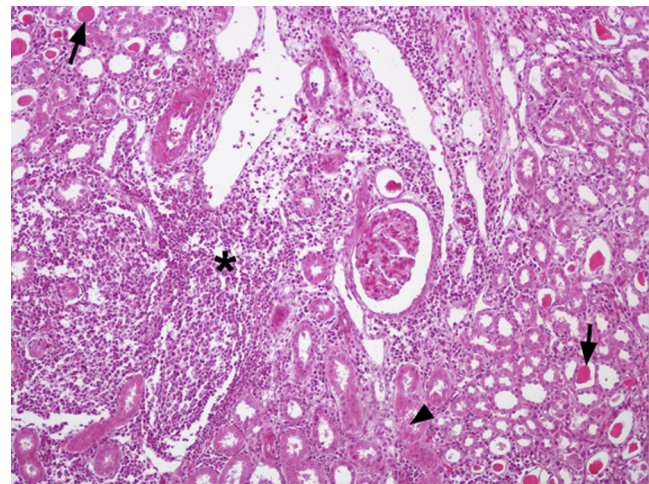


Figure 1 Kidney, multifocal infiltrations of mononuclear cells especially in cortical zone (*) and presence of eosinophilic material in proximal tubules are observed (→) as well as deposits of hyaline material in distal tubules and collecting duct (►). Staining haematoxylin-eosin. 200×.

Pomona, Pyrogenes, Ranarum, Celledoni, Hebdomadis, Sarmin, Bataviae, Autumnalis, Cynopteri, Panama, Australis, Mini, Javanica, Djasiman, Wolffi and Tarassovi.

Out of the 24 kidney samples, 10 had no histological lesions (they belonged to individuals with negative serological tests) and 14 kidneys had lesions which belonged to animals that presented specific antibodies to *Leptospira*, with serological titers from 50 to 3200 for serovars Canicola, Castellonis, Icterohaemorrhagiae and Hardjo. Animals which were serologically positive had a gradient of histopathological changes that were more severe in the animals with the highest titers. In the less severe cases the lesions found consisted of infiltrations of mononuclear cells, mostly lymphocytes and plasmatic cells around the renal corpuscles, whereas in the most severe cases the lesions had spread to the rest of the cortex, producing a cortical interstitial nephritis that could compromise the medullary area. In 9 of the 14 animals that tested positive there was a lymphocytic infiltration of plasmatic cells to the level of the submucosa in the renal pelvis. The parietal layer of Bowman's capsule was thicker in the most severe cases, with a retraction of the glomerular tuft and a dilation of the capsular space. In the renal tubules, especially in the collecting duct, there were cylindrical or teardrop-shaped deposits of hyaline material showing different levels of tubular degeneration (Fig. 1). Immunostaining was detected mainly inside the proximal tubules, added to the surface of epithelial cells, and between those cells in the three tested animals (Fig. 2).

The results of this study recognized positive serology of *Leptospira* in *C. villosus* in La Pampa province, where serovars Canicola (11.3%) and Castellonis (10%) were more prevalent and Hardjo (1.3%) and Icterohaemorrhagiae (1.3%) were less common. No significant differences of prevalence between males and females ($p=0.519$) were found; however, they were significant between juveniles and adults ($p=0.019$).

Cuba-Caparó⁴ identified serovars Hardjo, Wolffi, Sejroe, Hebdomadis, Bataviae and Canicola, with a prevalence

Table 1 Agglutination titers according to the MAT (microscopic agglutination test) of different *Leptospira* serovars reactants (Castellonis, Canicola, Hardjo, Icterohaemorrhagiae) in *C. villosus*, segregated by sex (F: female, M: male), age (adults, juvenile) and lesion presence.

Samples Adults	Sex	Serovar Castellonis	Serovar Canicola	Serovar Hardjo	Serovar Icterohaemorrhagiae	Histopathological lesions
609	F	50	50		50	n/p ^a
639	M		50			n/p
651	M		1600	100		YES
652	M		800			YES
653	F	3200	1600			n/p
660	F		100			YES
661	M	50	200			YES
664	M	100	800			YES
663	F			50		n/p
676	M		50			n/p
697	M	100			200	YES
698	M				50	n/p
703	M				50	YES
704	F	100	3200			YES
706	F	3200				YES
709	M		100			YES
727	M	400				n/p
739	M		800			n/p
742	F	50				n/p
773	F	50				n/p
776	F	100	100			n/p
786	F		800			n/p
788	M	400	800			n/p
789	M		200			n/p
791	F	400			200	YES
795	M	400				n/p
799	M	200				YES
800	F	800			200	YES
803	F	3200			800	n/p
810	F	1600	800			YES
815	F	800			200	n/p
825	M		800			n/p
836	M	1600			400	n/p
843	F	800			400	n/p
Juveniles						
744	f	50				n/p

^a Histopathology was not performed (n/p).

of 17.9%. Myers *et al*⁸. analyzed 89 *C. villosus* in Azul (Buenos Aires), resulting in 47.2% of animals that tested positive to different serovars including 1.1% for Pomona, 21.3% for Hardjo, Wolffi, Sejroe, Hebdomadis, 15.7% for Argentinensis, Paidjan and Bataviae, and 2.2% for Canicola; 6.7% of armadillos reacted to more than one serovar and antibodies against *L. biflexa* were found in one animal. These authors also mention the presence of serovar Argentinensis with 11.2%; however, this serovar could not be analyzed in armadillo sera from La Pampa because the strain is not currently available.

Scialfa *et al*¹⁰. analyzed five *C. villosus* from the province of Buenos Aires that tested negative to serovars Icterohaemorrhagiae, Canicola, Castellonis, Tarassovi, Pomona, Wolffi, Pyrogenes, Grippotyphosa, Hardjo and Hebdomadis, while another study in 2013¹¹ analyzed six *C.*

villosus, two of which tested positive to serovars Canicola, Castellonis, Grippotyphosa and Icterohaemorrhagiae (there was no discrimination of the serovars present in each animal). All samples were negative to serovars Tarassovi, Pomona, Wolffi, Pyrogenes, Hardjo and Hebdomadis.

Motie *et al*⁷. mention serovar Canicola for *Dasypus novemcinctus* in central Florida (EEUU) with a prevalence of 2.4% (7/86). The prevalence found by Myers *et al*⁸. (2.2%) was lower than the one found in this study (6%) for *C. villosus* in La Pampa.

The microscopic lesions observed in the kidneys of *C. villosus* agree with the ones described for leptospirosis in other armadillos^{8,12} and farm animals⁶. Detection of *Leptospira* in all tested animals confirmed the etiology of detected changes, and suggested that histopathology was useful for diagnosing the disease in the untested armadillos.

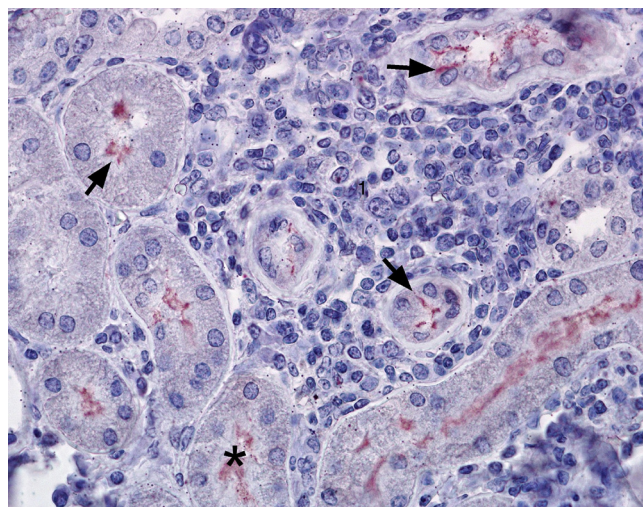


Figure 2 Kidney, immunostaining can be found as thin fibers between tubular cells and over the epithelial cells (arrows →), and with a diffuse pattern within the tubules (*). The interstitial tissue appeared infiltrated with lymphocytic cells (1). Immunohistochemistry, AEC (solution aminoethylcarbazole) – haematoxylin-eosin. 400×.

C. villosus adults presented the highest prevalence against *Leptospira* (26.7%) with respect to juveniles (4.3%). The difference could be a consequence of greater exposure time of adults to an environment contaminated with *Leptospira*.

Our results represent the first record of the presence of antibodies against *L. interrogans*, *Icterohaemorrhagiae*, *Canicola* and *Hardjo* serovars and antibodies against *L. borgpetersenii* Castellonis serovar for *C. villosus* in La Pampa, being the Castellonis and *Canicola* serovars those with the highest prevalence and the highest titers (3200). The presence of different serovars of pathogenic *Leptospira* in *C. villosus* shows a potential risk. Since *Leptospira* are mainly shed in urine, contaminating water, food and soil, new questions about other domestic and wild species should be answered. Furthermore, the importance of health education should be highlighted in order to raise awareness of the dangers involved in hunting, manipulating and ingesting armadillo meat, especially in areas where these habits are more frequent.

Ethical disclosures

Protection of human subjects

The authors declare that no experiments were performed on humans for this study. Experiments involving animals were performed respecting the guidelines of the Canadian Council of Animal Care, with the authorization of the Ministry of Production of La Pampa province, and with the permission of the farm owners.

Confidentiality of data. The authors declare that no patient data appear in this article.

Right to privacy and informed consent. The authors declare that no patient data appear in this article.

Conflict of interest

The authors declare that they have no conflicts of interest.

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References

1. Brihuega B, Tealdo M, Temas de zoonosis V. Capítulo 19. Importancia de los animales silvestres en la leptospirosis. AAZ; 2011. p. 169–74.
2. Cacchione RA, Cascelli E, Martínez ES, Zuberbuhler J. Leptospirosis en animales silvestres: aislamiento de una cepa de *Leptospira canicola* de un peludo (*Chaetophractus villosus*). Rev Med Vet. 1966;47:363–6 [On-line]. <http://www.sovergs.com.br/conbravet2008/anais/cd/resumos/R1179-2.pdf>
3. Carrizo A, Brihuega B, Etchechoury I, Arese A, Romero S, Gioffré A, Romano MI, Caimi K. Identificación de antígenos inmunorreactivos de *Leptospira interrogans*. Rev Argent Microbiol. 2009;41:129–33.
4. Cuba-Caparó A. The armadillo in biomedical research. In: Pan American Health Organization PAHO/ACMR 15/17. Fifteenth Meeting of the Advisory Committee on Medical Research. 1976. p. 1–43 [On-line]. http://hist.library.paho.org/English/ACHR/ACMR15_17.pdf
5. Delgado F, Capellino F, Venzano A, Funes D, Blanco Viera FJ, Auteri C, Romero G, Brihuega B. Adaptación de un protocolo de inmunohistoquímica para la detección de *Leptospira* spp. en muestras de tejido en formaldehído. Rev Cubana Med Trop. 2007;59:14–8.
6. Gardner AL. Orden Cingulata en Wilson y Reeder. Mammal species of the World. A Taxonomic and Geographic Reference. Third Edition Baltimore: The Johns Hopkins University Press; 2005. p. 94–103.
7. Jubb KVF, Kennedy PC. Patología de los Animales Domésticos. Agropecuaria Hemisferio Sur. Tomo II. Cap, vol. 6; 1980. p. 343–404.
8. Motie A, Myers DM, Storrs EE. A serologic survey for leptospires in nine-banded armadillos (*Dasypus novemcinctus* L.) in Florida. J Wildlife Dis. 1986;22:423–4.
9. Myers DM, Caparó AC, Moreno JP. Aislamiento del serotipo Hardjo y otras leptospirosis de armadillos de Argentina. Bol Sanit Panam. 1977;83:56–65 [On-line]. <http://hist.library.paho.org/spanish/Bol/v83n1p56.pdf>
10. Ojeda RA, Chillo V, Díaz Isenrath GB, editors. Libro rojo de mamíferos amenazados de la Argentina. Argentina: SAREM; 2012.
11. Scialfa EA, Brihuega B, Morris WE, Recavarren M, Quintana S, Grune S, Romero G, Bolpe J, Schettino M. First isolation of

- Leptospira interrogans* from *Conepatus chinga*. Afr J Appl Microbiol Res. 2012;1:1–5.
12. Scialfa E, Brihuega B, Venzano A, Morris WE, Bolpe J, Schettino M. First isolation of *Leptospira interrogans* from *Lycalopex griseus* (South American gray fox) in Argentina shows new MLVA genotype. J Wildlife Dis. 2013;49: 168–72.
 13. Stuart BP, Crowell WA, Adams WV, Carlisle JC. Spontaneous renal disease in Louisiana Armadillos (*Dasypus novemcintus*). J Wildlife Dis. 1977;13:240–4.
 14. Szyfres B, Sulzer CR, Galton MM. Nuevo serotipo de *Leptospira* del grupo *Bataviae* aislado en la Argentina. Bol Sanit Panam. 1968;64:225–7 <http://hist.library.paho.org/Spanish/BOL/v64n3p225.pdf>